Glioblastoma is the most common malignant brain tumor in adults. Even when aggressively treated with surgery, chemotherapy, radiation, and novel chemotherapies, the median duration of survival remains just shy of 2 years. While gradually improving over time, the prognosis for glioblastoma remains poor, and novel therapeutic approaches are desperately sought after.

Part of the lack of efficacy of chemotherapy drugs against glioblastoma results from the inability of most small molecules (98%) to pass the blood-brain barrier (BBB). The BBB usually prevents therapeutic agents with a molecular weight greater than 180 kDa from passing through. To overcome this, intraparenchymal delivery of therapeutic agents has been investigated. Approaches to local drug delivery have included the use of implantable polymers that slowly release drugs and convection-enhanced delivery (CED) through rigid cannulas and/or soft catheters. Here, we review the latter approach, including its conceptual origins, clinical trial results to date, current challenges, and future directions of the field.

Methods

A PubMed search was performed, using the phrase “convection-enhanced delivery and glioblastoma,” for all years up to 2015. The references of systematic reviews and preclinical or clinical studies in which CED was studied for the treatment of glioblastoma were reviewed based on survival outcomes, adverse effects, and methodologies. Studies focusing on physical mechanisms of CED were included, along with articles exploring technological CED advancements. Finally, studies elucidating the physiological properties and barriers in the brain related to drug delivery were used.

Results


Researchers seeking to bypass the BBB, which impedes many potentially effective treatments, by pursuing local delivery were initially limited to diffusion-mediated delivery. A compound’s diffusion in a given tissue depends mainly on its free concentration gradient and its diffusivity in the tissue. Thus, high-molecular-weight compounds like antibodies and enzymes are unable to diffuse over large distances, and drug distribution is very limited. For example, it may take up to 3 days for an immunoglobulin...
to diffuse 1 mm from its delivery site, which is far from ideal as a therapeutic method. 23

CED, initially conceptualized by Bobo et al. in 1994 as a local delivery strategy for the CNS, can overcome these limitations of diffusion-mediated delivery. 4 CED involves stereotactically inserting 1 or more rigid cannulas containing soft catheters into tumors through bur holes. The catheters are proximally connected to a syringe pump containing the infusate, while their distal ends protrude out beyond the tip of the cannula(s) into the targeted tumor area, establishing a positive pressure gradient for hours to days.

CED is powered by bulk flow kinetics from pressure gradients, as opposed to the concentration gradients relied upon for standard diffusion-based delivery. Although it was originally thought that infusates move through the interstitial space, there is evidence that molecules might move by other means, such as perivascularly, para-arterially, or through slower axonal transport. 27, 28, 39, 47, 61, 62

CED offers several advantages over diffusion-mediated delivery. First, CED improves intratumoral spatial distribution because the pressure gradient allows agents to be infused over a larger volume, more evenly and at higher quantities than in diffusion-based approaches (Fig. 1). 52, 72, 76 Second, because CED lacks the steep concentration gradients associated with diffusion-mediated delivery, less toxic doses can be used. 32 Third, unlike diffusion, CED occurs independent of an agent’s molecular weight or diffusivity.

Preclinical Development of CED for Glioblastoma: Summary of Select Studies

Since the 1994 study of Bobo et al., 4 CED for gliomas continues to be investigated in preclinical models. Here, we review various noteworthy preclinical studies. Intracerebral CED of topotecan in a rat glioma model demonstrated that 11 of 12 rats treated survived beyond 120 days while the control cohort did not live beyond 26 days. 25 Another study assessed CED of chemotherapies gemcitabine or carboplatin, in which 63% of animals receiving either agent via CED lived to the 120-day mark, while all control animals receiving phosphate-buffered saline via CED or systemic treatment expired before 26 days. 11 These studies, while promising, failed to compare CED of a therapy to systemic delivery of a drug with known efficacy. A study in 2004 did that by comparing baseline oral temozolomide (TMZ) efficacy to results achieved with the addition of TRAIL (tumor necrosis factor–related apoptosis-inducing ligand) via CED in athymic mice. This study established that the combination of TRAIL and TMZ prolonged overall survival (OS) compared with TMZ alone. 58 Moreover, a 2015 study revealed that bevacizumab, a humanized VEGF (vascular endothelial growth factor) antibody, administered via CED, increased survival over intravenous bevacizumab. This result supports further developing CED by showing that current glioblastoma treatments could be more effective via CED. 60 Other studies published in 2015 assessed the efficacy of CED of agents used to systemically treat glioblastoma such as irinotecan, carboplatin, and cetuximab delivered via nanoparticles and liposomes to allow gradual release after CED. 1, 26, 43 These preclinical studies have improved CED techniques and identified agents to infuse via CED, information that has given rise to the multiple clinical trials in patients described below.

Results of CED Clinical Trials 1997–2010

The results of CED clinical trials published to date are summarized in Table 1. Most trials have used flexible, single-lumen catheters rather than the rigid cannulas with tumor-penetrating catheters proposed in initial preclinical studies. 4 Of the 14 trials listed, 8 involved conjugated toxins specifically taken up by high-grade glioma cells, 1 involved viruses or liposomes, and 5 involved conventional chemotherapies unable to penetrate the BBB. For trials involving these chemotherapies, optimal cannula positioning safely away from the ventricles is vital to prevent leakage of...
<table>
<thead>
<tr>
<th>Authors &amp; Year</th>
<th>Agent Convected (type)</th>
<th>Trial Phase</th>
<th>Site of Study</th>
<th>Drug Classification</th>
<th>WHO Grade, Tumor Type (no. of pts)</th>
<th>Flow Rate</th>
<th>Vol Infused</th>
<th>Duration</th>
<th>Tracking</th>
<th>No. of Catheters</th>
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<tr>
<td>Laske et al., 1997</td>
<td>TF-CRM107 (synthetic protein exotoxin)</td>
<td>I</td>
<td>NIH/Temple</td>
<td>Synthetic protein endotoxin</td>
<td>Recurrent Gr IV GBM (10), Gr III AA (5), AO (1), metastatic lung CA (1)</td>
<td>Incrementally increasing w/ each Tx, up to 16 mg/day</td>
<td>5–180 ml</td>
<td>Up to 16 days</td>
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<td>Weaver et al., 2003</td>
<td>TF-CRM107 (synthetic protein exotoxin)</td>
<td>II</td>
<td>Temple</td>
<td>Synthetic protein endotoxin</td>
<td>Recurrent Gr IV GBM &amp; Gr III AA (44)†</td>
<td>0.2 ml/hr</td>
<td></td>
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<td>2</td>
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<tr>
<td>Rand et al., 2000</td>
<td>NBI-3001 (IL4-Pseudomonas exotoxin)</td>
<td>I</td>
<td>NIH</td>
<td>Chimeric recomb fusion protein</td>
<td>Recurrent or progr Gr IV GBM (9)</td>
<td>0.3–0.6 ml/hr</td>
<td>30–185 ml</td>
<td>4–8 days</td>
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<td>Weber et al., 2003</td>
<td>NBI-3001 (IL4-Pseudomonas exotoxin)</td>
<td>I</td>
<td>NIH</td>
<td>Chimeric recomb fusion protein</td>
<td>Recurrent Gr IV GBM (25), Gr III AA (31)</td>
<td>0.3 ml/hr</td>
<td>40–100 ml</td>
<td>1–45 days</td>
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<td>Lidar et al., 2004</td>
<td>Paclitaxel</td>
<td>III</td>
<td>Israel</td>
<td>Taxane chemo</td>
<td>Recurrent Gr IV GBM (13), Gr III AA (2)</td>
<td>0.18 ml/hr</td>
<td>4.5 ml</td>
<td>1–2 days</td>
<td>SPECT</td>
<td>1</td>
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<td>Patel et al., 2005</td>
<td>Cotara</td>
<td>III</td>
<td>4 diff'nt institutions</td>
<td>Chimeric recomb fusion protein</td>
<td>Recurrent Gr IV GBM (37), newly diagnosed Gr IV GBM (8), Gr III AA (6)</td>
<td>0.18 ml/hr</td>
<td></td>
<td></td>
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<td>Vogelbaum et al., 2007</td>
<td>IL-13 PE38QQR (CinTredekin besudotox)</td>
<td>I</td>
<td>NIH</td>
<td>Chimeric recomb fusion protein</td>
<td>Newly diagnosed Gr IV GBM (21), Gr III AO (1)</td>
<td>0.75 ml/hr</td>
<td>96 hrs</td>
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<td>Kunwar et al., 2007</td>
<td>IL-13 PE38QQR (CinTredekin besudotox)</td>
<td>I</td>
<td>NIH</td>
<td>Chimeric recomb fusion protein</td>
<td>Recurrent or progr Gr IV GBM (46), Gr III AA or AO (5)</td>
<td>0.75 ml/hr</td>
<td>96 hrs</td>
<td>Radiolabeled HAS</td>
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<tr>
<td>Kunwar et al., 2010</td>
<td>IL-13 PE38QQR (CinTredekin besudotox) vs Gliald wafers</td>
<td>III</td>
<td>52 diff'nt sites</td>
<td>Chimeric recomb fusion protein</td>
<td>Newly diagnosed Gr IV GBM (296)</td>
<td>0.75 ml/hr</td>
<td>96 hrs</td>
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<td>Sampson et al., 2008</td>
<td>TP-38</td>
<td>I</td>
<td>NIH</td>
<td>Chimeric recomb fusion protein</td>
<td>Recurrent or progr Gr IV GBM (20)</td>
<td>0.4 ml/hr</td>
<td>20 ml</td>
<td>50 hrs</td>
<td>Radiolabeled albumin</td>
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<td>Bogdahn et al., 2011</td>
<td>Trabedersen</td>
<td>II</td>
<td>NIH</td>
<td>Synthetic antisense phosphorothioate oligodeoxynucleotide</td>
<td>Recurrent or progr Gr IV GBM (103), Gr III AA (42)</td>
<td>4 μl/min</td>
<td>7 days</td>
<td></td>
<td></td>
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<tr>
<td>Bruce et al., 2011</td>
<td>Topotecan</td>
<td>I</td>
<td>Columbia</td>
<td>Cytotoxic quinoline alkaloid–derived chemo</td>
<td>Recurrent Gr IV GBM (10), Gr III glioma (6)</td>
<td>200 μl/hr</td>
<td>40 ml</td>
<td>100 hrs</td>
<td>Gadopentetate, gadodiamide, or gadobenate</td>
<td>2</td>
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<tr>
<td>Voges et al., 2003</td>
<td>HSV-1k</td>
<td>III</td>
<td>4 diff'nt institutions</td>
<td>Gene-bearing liposomal vector</td>
<td>Recurrent Gr IV GBM (8)</td>
<td>0.025–0.6 ml/hr</td>
<td>3.5 ml</td>
<td>29 days</td>
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</table>

AA = anaplastic astrocytoma; AO = anaplastic oligodendroglioma; chemo = chemotherapeutic agent; diff'nt = different; GBM = glioblastoma; Gr = grade; HSA = highly purified human serum albumin; lung CA = adenocarcinoma of lung; MAb = monoclonal antibody; mult = multiple; progr = progressive; pts = patients; recomb = recombinant; SPECT = single-photon emission computed tomography; Tx = treatment.

* Shown are published results of 13 clinical trials using CED in glioblastoma patients.
† Total of 44 patients; number per tumor type not specified.
chemotherapeutic agents into the CSF, which could cause aseptic chemical meningitis, although this complication has never been reported when chemotherapy is delivered into the parenchyma via CED. Below we summarize some important representative examples of these trials.

CED Trials Involving Conjugated Toxins

With promising results obtained from preclinical models, the first clinical trial was performed in glioblastoma using the targeted toxin TF-CRM107, a human transferin (TF) conjugated to diphtheria toxin (CRM107) with a point mutation that abrogates its nonspecific binding to mammalian cells. Results (reported in 1997) showed that 9 of 15 patients had tumor size reduction with limited toxicity, the first demonstration of CED feasibility in brain tumor patients. The Phase II arm for TF-CRM107, reported in 2003, produced slightly less encouraging results with only 31 of 44 patients completing treatment and only 12 of those 31 having a complete or partial radiographic response. Overall, 23 patients had progressive disease, while 21 patients had died; the median survival was 37 weeks. Toxicity included cerebral edema in 14% of patients. Unfortunately, the Phase III study involving TF-CRM107, conducted at 40 centers in the US and Europe was aborted because intermediate analysis of 44 patients showed only a 39% response rate. Temply reporting of aborted trials like the TF-CRM107 Phase III study and thorough analysis of potential sources of failure like cannula placement will be essential for the development of future CED trials.

Interleukin-4 (IL-4) receptor overexpression in malignant glioma cells was taken advantage of through the use of IL-4 Pseudomonas exotoxin (NBI-3001), a chimeric recombinant fusion protein made of IL-4 and Pseudomonas exotoxins. The first of such trials was in 2000 and involved a cohort of 9 patients with glioblastoma, 6 of whom demonstrated tumor necrosis after treatment, a promising response with limited toxicity. In a similar NBI-3001 trial, a cohort of 6 Grade III astrocytoma patients and 25 glioblastoma patients received CED as their only treatment. The median OS for this study was 8.2 months; patients with glioblastoma had a median OS of 5.8 months. Unfortunately, the Phase III study involving TF-CRM107 demonstrated a significant amount of tumor necrosis in a majority of these patients. There was no systemic toxicity, and adverse effects were limited to treatable edema. An additional case study involving a patient with recurrent glioblastoma treated with NBI-3001 in a single infusion showed that the patient survived for 36 months. The IL-4–Pseudomonas exotoxin fusion protein holds promise as a targeted therapeutic for glioblastoma and is being developed by Medicenna Therapeutics.

Epidermal growth factor receptor (EGFR) overexpression is another feature of glioblastoma that can be therapeutically targeted. TP-38 is a chimeric protein containing a Pseudomonas exotoxin and a TGF-α binding domain that binds tightly to EGFR. Its binding delivers the exotoxin and induces apoptosis. In a study of 20 patients with recurrent or progressive malignant brain tumors (17 glioblastomas), the median time to progression was 14.9 weeks, and the median OS was 28 weeks. However, many patients experienced significant leaks into the ventricles or subarachnoid space resulting in failed intraparenchymal distribution. Although the treatment was considered safe, this trial revealed the importance of real-time CED monitoring. In a case study involving a patient with multiply recurrent glioblastoma, TP-38 prevented tumor progression for over 43 months, with PET revealing hypometabolic tumor activity, suggesting treatment response.

Beyond the Phase III TF-CRM107 study mentioned above, the only other randomized Phase III CED clinical trial to date studied IL13-PE38QQR (Cintredekin besudotox), a chimeric protein made up of IL-13 and a truncated form of Pseudomonas exotoxin A (PE38QQR), which is taken up by glioma cells expressing the IL-13 receptor alpha 2 chain (IL-13Rα2). This trial (named the PRECISE Trial) enrolled 296 patients; one arm received IL13-PE38QQR via CED through catheters implanted after craniotomy for resection 96 hours earlier, and the other received CCNU-containing Gliadel wafers implanted in the walls of the resection cavity after craniotomy. Although no OS benefit was found for CED of IL13-PE38QQR relative to Gliadel wafers, the study was powered to detect greater than 50% survival benefit over the Gliadel-treated control arm, and detecting a smaller but meaningful difference would have required a larger cohort, significantly elevating the trial’s cost. A follow-up study noted that only 68% of catheter placements were performed per protocol, suggesting that variability in catheter positioning may have adversely impacted results. The authors also commented that excluding patients with lower IL-13 receptor levels might have improved efficacy. Despite failure to improve OS, the prespecified objective for efficacy, improved progression-free survival (PFS) (18 vs 11 weeks, p = 0.0008) was reported.

CED Trials Involving Conventional Chemotherapy

Paclitaxel, a chemotherapeutic agent unable to cross the BBB, was studied in a CED trial for the treatment of high-grade gliomas published in 2004 by Lidar et al. Thirteen glioblastoma and 2 anaplastic astrocytoma patients with recurrent disease received a total of 20 cycles of paclitaxel via CED. Eleven of 15 patients showed an imaging response, and the median survival was 7.5 months. Camptothecin class drugs inhibit topoisomerase I, thereby disrupting DNA replication and triggering apoptosis. Topotecan, a member of this class, was used in a CED trial with 16 patients (10 glioblastoma, 6 anaplastic astrocytoma). The trial’s median OS and PFS were 60 and 23 weeks, respectively. Topotecan was not detected in patient serum, and patients receiving the highest dose suffered only minor neurological deficits.

CED Trials Involving Liposomes or Viruses

Implanted liposomes, spherically shaped artificial vesicles bearing a lamellar phase lipid bilayer, can be used for gradual therapeutic delivery. Nanoparticles within the liposomes can be modified to control drug release and improve target-site specificity. Liposomes can also undergo endocytosis or phagocytosis, allowing intracellular delivery of drugs that cannot pass across the cell membrane. Liposomes have been used to deliver genes to tumor cells in several trials. Initial attempts at gene delivery to
tumor cells involved implanting vector-producing cells generating retroviruses carrying the herpes simplex virus–thymidine kinase (HSV-TK) gene. After these attempts failed to produce sufficient tumor cell transduction, intratumoral delivery of nonreplicating adenoviruses containing HSV-TK without convection was studied in clinical trials. The lack of distribution sufficiently beyond the injection site in these studies created an interest in HSV-TK gene delivery via CED. In 2003, Voges et al. reported results from a trial in which the HSV-TK gene in liposomes was delivered to glioblastomas via CED. This study involved 8 patients, who had a median survival of 28.1 weeks, with no morbidity witnessed from the procedure. Two of 8 patients had tumor reduction over 50%, although the distribution volume was small in this trial. Another study used liposomes carrying nonreplicating Semliki Forest virus with IL-12, an agent that activates natural killer cells, although the results were inconclusive. Further evaluation of liposomal delivery of therapeutics via CED is clearly warranted.

**Challenges Associated With CED**

Although CED holds considerable promise in neurological drug delivery, clinical trials have had limited success. Reasons identified for these failures are summarized below.

**Choice of Agent for CED**

While considerable attention has been given to technological development of CED (e.g., cannula design and infusion rates), the choice of agent to be delivered is exceedingly important in dictating the success of CED. In particular, the inclination to use particularly toxic agents for CED based on the presumption that local delivery ensures the agent only gets to the tumor must be avoided. This is because of the need to target not just the tumor but also the infiltrating tumor cells within the peritumoral white matter, which also contains normal brain cells. Thus the delivered agent must possess a wide therapeutic index. Many CED trials have therefore employed molecularly targeted agents whose toxicity is limited to tumor cells. As is the case for systemically administered agents, Phase 0 clinical trials may be needed to narrow the large number of agents being considered for CED. In Phase 0 trials, subtherapeutic doses of an agent are given to a small number of patients, followed by tissue procurement to confirm that the agent’s target has been inhibited. For CED, this would involve a craniotomy after CED to confirm achievement of 2 CED goals: target inhibition by the drug and sufficient intratumoral distribution of the drug by CED.

**Cannula Design**

Several improvements in cannula design made over the original rigid cannulas together allow for increased infusion rate, reduced reflux, and increased infusate distribution. Rounded-tip cannulas, for example, were introduced to decrease tissue pressure and thereby reduce tissue damage–related reflux. The step cannula (Fig. 2) has a distal tip that is smaller in diameter than the rest of the cannula; this design offers the maneuverability of large-diameter cannulas with the higher infusion rates afforded by small-diameter cannulas. A single patient study using a recessed-step cannula (step cannula with outer reinforcement) and robot-guided infusions reported 95% tumor volume coverage.

Multiple modifications have been attempted to improve flow from the cannula. Cannulas were designed with multiple catheters at their tips, which would have lowered infusion rates at each opening. Unfortunately they were ineffective, since most of the infusate was distributed via the proximal tip. Hollow-tipped cannulas with multiple nano-sized openings for infusate delivery showed in a murine model 3 times higher distribution of infusate compared with single opening cannulas. Additionally, the balloon cannula (Fig. 2) allows for a large amount of surface area contact within the tumor, confirmed by a canine model showing 90% infusate distribution with its use.

Longer-term CED will also benefit from cannulas with flexible catheters (Fig. 2), which have been used in most clinical trials to date (Table 1). These cannulas are rigid during insertion due to their rigid interior stylet but become flexible after they have been inserted and the stylet has been removed. These cannulas could allow long infusions in mobile patients, and implantation for up to 5 days has been approved. Cannulas composed of biocompatible materials such as carbothane have been shown to be suitable for even longer-term insertion, with homogenous distribution of infusates and adequate delivery rates (5 μl/min) without reflux or tissue adherence.

**Cannula Placement**

Retrospective modeled distribution analysis of the PRECISE trial suggested that less than 50% of the patients had optimal cannula positioning, which could explain the lack of survival benefit in the trial, given that there was a positive correlation between longer survival and cannula placement considered ideal on the basis of modeling. Although there are numerous assumptions in this analysis, further study and modeling of optimal cannula positioning is clearly warranted. Concepts related to optimal cannula positioning that are fully validated should be incorporated into future CED clinical trials, including concepts lacking in the trials summarized above, such as software algorithms for cannula positioning, mandating formal training in cannula positioning for neurosurgeons participating in these trials, and identifying whether there is a neurosurgical learning curve for cannula positioning that should be accounted for by mandating that neurosurgeons participating in Phase III CED trials have prior experience with Phase I and II CED trials to ensure that Phase III trials are adequately able to assess efficacy. These stringent criteria may initially limit the generalizability of CED and restrict it to specific providers or specific centers, but demonstrated efficacy of CED in a trial designed with these considerations in mind would likely over time increase the commitment of other centers and providers to obtain the platforms and technical skill needed to implement CED.

**Intratumoral Penetration**

The heterogeneous nature of tumors makes homogeneous delivery of therapeutic agents challenging. Several
physiological and physical barriers prevent full tumor distribution. One physiological barrier is that certain tumor zones metabolize agents faster than others, while a physical barrier can be the aberrant blood vessel growth and increased intercapillary spaces that create differential rates of clearance of agents. Modification of therapeutic agents can improve intratumoral penetration by CED by allowing accurate predictions of concentration distribution.

Reflux
A major concern with CED is infusate reflux, which has been noted in several trials. Reflux occurs when the pressure gradient between the cannula and the tumor region equalizes, resulting in the loss of drug flow into the target mass. Three variables have been suggested to contribute to reflux: tissue disruption at the tip of the cannula, infusion rate, and cannula diameter. Tissue disruption causes reflux when a cannula is inserted too deeply into tissue, if there is a brain shift after the cannula has already been inserted in the brain, or if biopsies are taken at the cannula tip prior to infusing. Tissue disruption potentiates formation and subsequent filling of a cavity, creating a pressure reservoir that ultimately causes backflow up the catheter and outside its wall. Softer cannulas and catheters (Fig. 2) and quick catheter placement reduce brain shift–induced tissue disruption.

High infusion rates increase reflux risk because pressure caused by the infusion process may exceed interstitial pressure sufficiently to promote reflux. Fortunately, modern cannula design described below has allowed infusion rates up to 50 μl/min without reflux. However, infusion rates of 3 μl/min or higher can drive infusate into the subarachnoid and ventricular space, causing adverse effects—underscoring the importance of safe target selection. In designing CED protocols, infusion rate and cannula diameter need to be considered together as they interact such that each cannula diameter and flow rate will be associated with a specific predicted backflow distance along the cannula.

Tracking Infusate Delivery
None of the trials described above assessed the efficacy of delivery. Two preclinical studies, case reports of 1 or 2 patients, and more recent clinical trials whose results have not yet been published have tracked infusate delivery by mixing MRI contrast agents like gadolinium-DPTA and gadoteridol-loaded liposomes (GDL) with infusate and using real-time intraoperative imaging in a surgical MRI suite (NCT02022644 and NCT01156584, ClinicalTrials.gov). This approach has 3 benefits: 1) validation of accurate cannula placement; 2) tracking of agent distribution within the tumor to allow identification of leakage as was done with GDL in an animal study; and 3) real-time adjustments, including choice of extra targets.

Tracking delivery by mixing the therapeutic agent with a contrast agent assumes that the two agents spread through the tumor similarly. Linking drug to a molecule that can be imaged, assuming the linkage does not disrupt drug efficacy, would address this issue. When patients cannot tolerate gadolinium, T2-weighted imaging can track infusate delivery, although glioblastomas often start with T2 hyperintense regions that would preclude such an approach. Validation of these methods for tracking infusate delivery may eventually lead to mandating these measures in Phase III trials.

Cost of Procedure
For the moment, the cost of CED, which includes time
Convection-enhanced delivery in glioblastoma

in the operating room and/or MRI, cannulas and the platforms needed to place them, infusates, time of the staff performing the procedure, and associated inpatient stays, may limit its applicability to smaller Phase I trials and Phase III trials like PRECISE, for which the corporate sponsor had sufficient funds to conduct an appropriately powered study. But, as with any medical innovation, procedural costs will decline as more companies enter the field and as innovation increases cost-efficiency. For example, the ClearPoint navigation platform (MRI interventions), which is commonly used in CED, began with a skull mount frame requiring skull exposure comparable to craniotomy, but recently a scalp mount frame has been developed, allowing the cannula to be passed through a nick in the skin, saving an hour of procedural time and reducing infection risk and cost.

Attaching CED to other procedures can allow for insurance coverage of operating room time and hospital stay, but only if that procedure is performed at the same time as CED, which is challenging, because a biopsy, while helpful for confirming tumor recurrence at the site of CED, can create blood products that adversely impact CED, and a craniotomy immediately before CED, although used in the PRECISE trial, creates a cavity, which allows reflux.

Postprocedural Imaging

The optimal endpoint for CED remains unclear. Until we develop ports that can be implanted long-term through which patients can receive multiple rounds of CED over a prolonged period of time, CED will be a one-time treatment, with treatment durability dependent on the agent being convected—lower for conventional chemotherapy but higher for chemotherapeutic agents in liposomes or replicating viruses. Thus, short-term imaging response such as reduced enhancement or FLAIR may be needed as a response metric alongside conventional parameters, such as PFS or OS, for these trials. The fact that IL13-PE38QQR affected PFS but not OS in the PRECISE trial underscores the fact that short-term metrics may better reflect CED success until technology renders multiple CED treatments per patient more feasible. Allowing multiple treatments may enable CED to impact conventional metrics like OS, because unlike agents like bevacizumab, which similarly impact PFS but not OS, the impact of CED of an agent like IL13-PE38QQR on PFS rather than OS may reflect reduced intratumoral persistence of the agent rather than the tumor evolving resistance to the therapy.

Future Directions

Here, we have outlined numerous technical challenges that need to be met to overcome the issues encountered with the use of CED to treat glioblastoma to date. Another consideration that will be important to prioritize going forward is that durable CED efficacy might require long-term convection at set intervals for months, as is often required for systemically administered chemotherapy to be effective for nonbrain tumors. The success of such a strategy may require implantable ports that can be cannulated to receive CED in an outpatient setting. Finally, before we can assess the efficacy of an agent delivered via CED, technical reproducibility must be achieved.

Conclusions

CED bypasses the BBB and reduces systemic toxicity. It has several advantages compared with diffusion-mediated delivery, as pressure gradients allow agents to be disturbed over larger volumes and more evenly at higher quantities, and its distribution is not limited by the drug’s physical characteristics. Thus, antitumor drugs that are often toxic systemically can be delivered at higher concentrations with excellent intratumoral distribution. Several therapies have been shown to be safe and somewhat effective in preclinical and clinical studies. Discouraging results from the two randomized Phase III trials conducted to date revealed technical shortcomings that need to be addressed to allow CED to fulfill its therapeutic potential. Overall, CED holds promise for treating glioblastoma and warrants further preclinical and clinical development.

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Disclosures
The authors report no conflict of interest concerning the materials or methods used in this study or the findings specified in this paper.

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Conception and design: Aghi, Jahangiri. Acquisition of data: Jahangiri, Chen. Analysis and interpretation of data: Jahangiri, Chin. Drafting the article: Jahangiri, Chin. Critically revising the article: Aghi, Flanigan, Chen, Bankiewicz. Reviewed submitted version of manuscript: Aghi, Jahangiri, Flanigan. Approved the final version of the manuscript on behalf of all authors: Aghi.

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