The use of fluorescence imaging technologies in neurosurgery has been widely reported dating back to 1948.\textsuperscript{13} Although the first reports of fluorescence angiography to visualize cerebral vessels using fluorescein appeared in 1967, recent reports have focused on indocyanine green (ICG) due to the development of a practical microscope-integrated ICG-imaging infrared module.\textsuperscript{3} Indocyanine green fluorescence angiography has been a useful adjunct in aneurysm surgery, as it allows for timely and safe evaluation of large vessels, aneurysms, and small perforating arteries exposed at surgery. For these reasons, it has also been evaluated as an adjunct in the microsurgical treatment of arteriovenous malformations (AVMs).\textsuperscript{8} Takagi et al.\textsuperscript{25} noted that ICG is easier to detect and characterized by a lower rate of adverse reactions than fluorescein. However, the recent development of a microscope-integrated fluorescein videoangiogram module allows for administration of small amounts of fluorescein with the added ability to visualize the operative field under a real-time fluorescence mode.\textsuperscript{10,11,19,20} These new advancements offer easy intraoperative detection of fluorescein and an improved rate of adverse reactions, which, based on ophthalmological studies, are comparable to those of ICG.\textsuperscript{4,29}

In this study, we report on the use of this newly developed microscope-integrated fluorescein videoangiogram module in the evaluation and treatment of AVMs. To our knowledge, this is the first report on the use of this technology for AVM surgery.

**Methods**

**Patient Population**

This study includes prospective data on 4 consecutive patients in whom AVM resection was performed using...
Sodium Fluorescein Dye

Sodium fluorescein has been used extensively in ophthalmological angiography of the retina. It is commercially available as a small organic molecule supplied in an aqueous solution with a pH of 8.0–9.8 and osmolality of 727–858 mOsm/kg. Sodium fluorescein fluoresces with wavelengths of 520–530 nm in response to excitation with light at 465–490 nm wavelengths. In practice, this results in yellowish-green fluorescence that can be detected by videoangiography or by the observer’s naked eye, at low and high intravenous doses, respectively.

Pharmacokinetically, fluorescein distributes well into the interstitial space, with an estimated volume of distribution of 0.5 L/kg. Approximately 20–30 seconds after a bolus peripheral venous administration, fluorescein can be observed in the cerebral circulation.

Fluorescein metabolism is primarily achieved through conversion to fluorescein monoglucuronide in the liver. This process is rapid, with approximately 80% conversion within 1 hour of injection. Furthermore, fluorescein is excreted via renal clearance at a rate of 1.75 ml/min/kg. Systemic clearance ranges from 48 to 72 hours in doses of 500 mg used in ophthalmological studies. Fluorescein contraindications only include those with known hypersensitivity to sodium fluorescein or other ingredients in the clinical solution (http://dailymed.nlm.nih.gov/dailymed/lookup.cfm?setid=ebb3883c-71f6-4fd0-a7e6-0ba8e1136dd9).

Due to its popularity in ophthalmological angiography, fluorescein’s side effect profile has been well characterized. Yanuzzi et al. performed a large complication survey in 1984 that included 2434 ophthalmologists and more than 220,000 angiograms. Side effects were categorized as mild, moderate, or severe. Mild and moderate reactions were classified both as transient in effect and with no subsequent sequelae or threat to patient safety. Specifically, mild reactions included nausea, vomiting, sneezing, pruritis, extravasation, and inadvertent arterial injection. Approximately 87% of survey respondents indicated a frequency of mild reactions of less than 5%. Moderate reactions included urticaria, syncope, other skin eruptions, thrombophlebitis, pyrexia, local tissue necrosis, and nerve palsy. These were reported with a frequency rate of 1 per 63 cases. Severe reactions were defined as those involving prolonged effects requiring intense treatment and resulting in variable recovery. These included laryngeal edema, bronchospasm, anaphylaxis, circulatory shock, myocardial infarction, cardiac arrest, and tonic-clonic seizure. These reactions were reported at a rate of 1 per 1900 cases. One death was reported in the study (rate of 1 per 222,000 cases).

High-dose fluorescein (20 mg/kg) has been used in neurosurgery for fluorescence-guided glioma resection, and there have been 2 reports of anaphylaxis with this large dose. It is important to note that these studies reported only on the use of high-dose fluorescein. Fluorescein does not cross the intact blood-brain barrier and remains largely intravascular. Extravascular fluorescein leakage is due to loss of integrity of the blood-brain barrier. Dural vessels stain as a result of this phenomenon and can interfere with imaging of neighboring structures. Large vessels tend to stain with repeated use. This is likely due to accumulation of fluorescein within the vessel wall; we have not observed this phenomenon with smaller vessels.

Microscope-Integrated Fluorescein Module

We performed intraoperative videoangiography using an OPMI PENTERO 900 operating microscope (Zeiss Meditec), which is equipped with the YELLOW 560 microscope-integrated module. This module utilizes stimulus light between 400 and 500 nm and detects wavelengths in the range of 540 to 690 nm for display of fluorescein. These specifications match those of sodium fluorescein fluorescence. This module has recently been integrated into the surgical microscope to allow the surgeon the ability to switch between stimulus light and white-light illumination by merely pressing a button. The optics of this module allow for visualization of nonfluorescent structures in nearly natural hue under fluorescent mode imaging. Furthermore, the module allows for high-definition digital recording of the operative field and fluorescent structures. This combination maximizes tissue details and optimizes microsurgical work while imaging in fluorescent mode.

The INFRARED 800 fluorescence camera, used for ICG imaging, separates emission and excitation light. Therefore, only fluorescent areas are visible in standard definition quality, but nonfluorescent areas appear black. The INFRARED 800 is designed for an excitation range of 700–780 nm and emission detection in the range of 820–900 nm. The ICG signal emission cannot be seen through the operating oculars and must be processed and displayed on a monitor. This technology does not allow for visualization of operative field under the fluorescent mode. The phenomenon of chromatic aberration significantly degrades image quality under high magnification of the operating microscope.

Technique for Fluorescence Angiography

After craniotomy and exposure of the AVM, a 75-mg

<table>
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<th>Case No.</th>
<th>Age (yrs)</th>
<th>AVM Location</th>
<th>Size (cm)</th>
<th>Preop Embolization</th>
<th>Presentation</th>
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<td>3</td>
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</tr>
<tr>
<td>4</td>
<td>33, M</td>
<td>rt frontal</td>
<td>2</td>
<td>no</td>
<td>seizures</td>
</tr>
</tbody>
</table>
Fluorescein videoangiography

bolus of sodium fluorescein (Akorn, Inc.) was administered through a peripheral intravenous line, and the fluorescein module was used for imaging. Fluorescent signal was apparent through the operating oculars after 10–15 seconds within the cortical vessels. This small dose is 15% of the dose previously used for neoplastic and vascular applications.3,5,9,15,22,24 We chose this dose based on our previous experience. We had decreased fluorescein dosing gradually from 200 mg in 25 increments for the previous 10 patients who harbored brain aneurysms. Based on this experience, we concluded that 75 mg was the lowest dosage that provided good fluorescent vascular imaging.

We used the following steps to perform 2–3 intraoperative videoangiograms for each AVM. The first videoangiogram was completed before AVM resection and evaluated normal cortical versus AVM feeding arteries (angioarchitecture). It also delineated the estimated cortical boundaries of the AVMs.

After most of the feeding arteries were disconnected, the status of the draining veins was assessed before their disconnection, using additional videoangiograms. More specifically, 1–2 additional injections (the number being based on the number of large draining veins present for the AVM) of fluorescein (75 mg dosing repeated) were used to assess the significance of draining veins for the partially disconnected AVM. The large draining veins with slowest flow as demonstrated through videoangiography were sacrificed first. The disconnection of this cortical vein allowed further mobilization of the AVM and facilitated further AVM disconnection until the AVM was completely excised.

Results

All 4 patients underwent fluorescein angiography with no complication. In each case, videoangiography allowed a better understanding of the AVM’s angioarchitecture and recognition of feeding arteries and draining veins. In one case involving a large frontal AVM, videoangiography demonstrated mainly cortical veins on the surface of the AVM and alerted the senior author (A.A.C.-G.) to first tackle the feeding arteries in the interhemispheric space. Videoangiography also prioritized the order for disconnection of large draining veins to allow mobilization of the AVM and exposure of the deep arterial feeding vessels.

In the other 3 cases, videoangiography readily allowed recognition of the AVM angioarchitecture and estimation of its cortical boundaries and, most importantly, assessed the flow within the draining veins before their disconnection. The ability to visualize the above fluorescence information through the operating oculars allowed the operator to readily appreciate the significance of this information in relation to the surrounding normal neurovascular structures. Ultimately, assessment of flow within the draining veins to prioritize their disconnection was the most important contribution of videoangiography. Postresection videoangiography was not performed as potential small deep residual AVMs are often not in the field of view of the microscope due to coverage by the surrounding cortex and white matter. Only postresection intraoperative arteriography was used to confirm a complete excision of the AVM.

Illustrative Cases

Case 1

A 35-year-old man presented after a seizure and was found to harbor a left frontal 4-cm AVM. Following embolization, he underwent microsurgical resection. After a frontal craniotomy and opening of the dura (Fig. 1A), fluorescein videoangiography revealed the cortical surface of the AVM. Based on this imaging, the surface cortical vessels associated with the AVM were determined to be predominantly draining veins (Fig. 1B). Although our initial plan was to disconnect the surface arterial feeders and then tackle the interhemispheric feeders originating from anterior cerebral arteries, we changed our plan based on fluorescein angiography information and first pursued the disconnection of interhemispheric feeders.

Following disconnection of the majority of superficial feeders, including the interhemispheric ones, fluorescein videoangiography was performed again to assess the flow status within the 2 draining veins in order to determine which vein could most safely be sacrificed to allow mobilization of the AVM and handling of deeper white matter feeders. This second fluorescein videoangiogram showed that the flow within the anterior vein was less that that in the posterior one (Fig. 1C), and therefore the former was disconnected, allowing the AVM to be mobilized posteriorly for its final excision.

Case 2

A 32-year-old woman presented with intractable headaches and was diagnosed with a 3-cm left posterior parietal AVM. Following its embolization, the AVM was exposed through a parietal craniotomy (Fig. 2A). The initial videoangiogram demonstrated the angioarchitecture of the AVM (Fig. 2B). Following disconnection of the superficial feeders, another videoangiogram was performed (Fig. 2C); this revealed minimal flow within the parasagittal vein, which was disconnected to mobilize the AVM and allow for its complete excision.

Discussion

Fluorescence angiography is a recent development that uses both fluorescein and ICG as fluorophores to image cerebral vessels.5,10,17,21,23,28 Fluorescein and ICG carry many of the same advantages and disadvantages. Both are easy to administer, inexpensive, and require no extra equipment or personnel. Fluorescein and ICG are equally effective in illuminating AVM vessels, including cortical feeding arteries and draining veins, as compared with digital subtraction angiography (DSA). Unfortunately, both ICG and fluorescein are only useful at visualizing vessels within the field of view of the microscope. This fact is a major limitation for confirmation of complete AVM resection, and therefore DSA remains the gold-standard technique for this purpose.

Although ICG and fluorescein have many similarities, they are characterized by unique chemical properties,
which confer distinct advantages and disadvantages. The combination of fluorescein and the YELLOW 560 module allows the surgeon to evaluate the surgical field in real time while looking through the microscope oculars under the fluorescent mode. This is possible since the emission of fluorescein falls within the visible electromagnetic spectrum, allowing for visualization of nonfluorescent structures in near-natural colors. This phenomenon allows the surgeon to mobilize and manipulate structures and more readily assess the fluorescent signal in relation to the surrounding cerebrovascular structures. As previously noted, repeated administration of fluorescein tends to stain the vessel walls in large caliber vasculature.18 We did not observe this effect using the smaller dosing of fluorescein for our repeated injections for flow assessment within the draining veins.

Indocyanine green has been studied by multiple groups, and has been noted to easily provide relevant information integrated into a separate monitor.2,6,8,14,25 Major reported advantages of ICG include its rapid clearance and therefore its application for multiple injections. In addition, the FLOW 800 module allows objective time-dependent assessment of intraoperative blood flow during removal of AVMs.2,6,14 Unfortunately, ICG imaging using the INFRARED 800 module separates fluorescent and nonfluorescent signals, whereby only the fluorescent signal is visible through a designated monitor and all other signals are muted to black. It has been our experience that this phenomenon prevents safe manipulation of structures in the operative field under the fluorescent-mode microscopy.

As fluorescence angiography with either fluorescein or ICG is insufficient to detect feeders or residual AVM beneath the brain parenchyma or outside the microscope field of view, DSA remains the ideal surgical adjunct to detect residual deep-seated AVM. However, DSA is also associated with a number of drawbacks, including associated morbidity, expense, increased operating time, need for additional resources and personnel, and added interpretation of study results, because the information is not integrated into the surgeon’s viewing field.8 Some have advocated for the use of Doppler ultrasonography to detect vessels associated with deep lesions beneath the brain parenchyma, but this modality lacks high resolution and does not offer a suitable alternative.12,27

Conclusions

Fluorescein and ICG have complementary advantages and disadvantages as fluorescent agents, and the use of fluorescein may provide additional advantages in the surgical management of select AVMs. Fluorescein

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Fluorescein videoangiography

videoangiography allows the surgeon to manipulate the surgical field under the fluorescent mode microscopy. We conclude that intraoperative fluorescein videoangiography can be potentially useful in the treatment of AVMs. This is a preliminary study, and larger studies are necessary to better understand the efficacy of and indications for this technology.

Disclosure

At the time this study was conducted and written, the authors reported no conflict of interest concerning the materials or methods used in the study or the findings specified in this paper. After this study was accepted for publication, Dr. Cohen-Gadol agreed to a consulting contract with Carl Zeiss Meditec. The compensation from this contract is donated entirely to a not-for-profit educational organization, The Neurosurgical Atlas.

Author contributions to the study and manuscript preparation include the following. Conception and design: both authors. Acquisition of data: both authors. Analysis and interpretation of data: both authors. Drafting the article: both authors. Critically revising the article: both authors. Reviewed submitted version of manuscript: both authors. Approved the final version of the manuscript on behalf of both authors: Cohen-Gadol.

References


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