Genetic investigation of multicentric glioblastoma multiforme: case report

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The authors report a case of multicentric glioblastoma multiforme (GBM) in which all 4 tumor foci were resected and evaluated using both comparative genomic hybridization array and RNA sequencing. Genetic analysis showed that the tumors shared a common origin, although each had its own unique set of genetic aberrations. The authors note that the genetic heterogeneity of multicentric GBM likely contributes to the failures of current treatments. The case underscores the necessity of increased genetic investigation.

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Glioblastoma multiforme (GBM), a Grade IV astrocytoma, is the most malignant and most common primary brain tumor.8 Recent studies suggest that nearly 80% of all malignant brain tumors are gliomas, and 54% of all malignant brain tumors are GBMs.4 The standard of care for GBM is resection followed by adjuvant radiation and chemotherapy, yet median survival times remain poor.14 The prognosis is worse for patients with multiple lesions than for those with a single focus— with a median overall survival of 10 versus 18 months in one recent single-institution retrospective study.15 Systematic studies analyzing GBM genomics demonstrate substantial intertumoral heterogeneity.2,3,10 There is also increasing evidence to suggest that GBM tumors have considerable intratumoral heterogeneity and are made up of genetically unique subclones that can evade standard therapy modalities, resulting in disease progression.5,9 Furthermore, a recent study found a high level of cell-to-cell diversity within individual GBM tumors.11

Case Report

History and Examination

This 47-year-old man presented to an outside clinic with leg weakness, near falls, and left inferior quadrantanopia, as well as progressive right-sided headache that primarily occurred upon standing. His medical history was significant for a lumbar laminectomy for spinal stenosis. The initial workup included a CT scan, which revealed 3 right-hemisphere lesions, leading to the patient’s transfer to our neurosurgical clinic. Neurological examination revealed no confusion, no problems with memory, and no language-associated problems. Based on these findings, a brain MRI was ordered.

On MRI, 3 enhancing mass lesions were identified along with hippocampal involvement (Fig. 1). The masses were in the right frontal lobe (2.6 × 2.5 × 4.4 cm), right occipital horn region (2.1 × 3.6 × 2.0 cm), and right temporal lobe (2.7 × 4.1 × 2.9 cm). The relative cerebral blood volume (rCBV) was variably elevated for these lesions, measuring 3.1, 3.0, and 3.1 times that of normal-appearing, contralateral white matter, respectively, as measured by first-pass dynamic contrast enhancement (DCE). Each of these lesions demonstrated marked surrounding hyperintensity on FLAIR sequences. Additionally, there was a fourth non-enhancing, FLAIR-hyperintense mass lesion involving the right hippocampus (rCBV = 2.1). The FLAIR sequences
showed that the lesion in the right frontal lobe was well separated from the others. FLAIR imaging did not show evidence of tracts connecting the tumors, and enhancing lesions had hyperintensity that radiated away from them (infiltrative edema) with minimal overlap. The appearance was not one of unidirectional extension of FLAIR hyperintensity with multifocal enhancing components, or gliomatosis cerebri.

Operation

The patient underwent craniotomies for removal of tumors in the right frontal lobe, right occipital horn region, right temporal lobe, and hippocampus. Intraoperatively, all of the tumors appeared grossly abnormal—grayish and hypervascular—and complete resections were achieved. Specimens were sent to pathology with frozen sections confirming GBM for each lesion. The remaining samples were sent to the laboratory for genetic analysis and cryogenic preservation.

Histological Evaluation

Histopathological examination (Fig. 2) of each sample using H&E stain showed brain tissue with a highly cellular pleomorphic astrocytic neoplasm. Microvascular proliferation and significant necrosis was present in each sample except for those from the hippocampal lesion. The histological diagnosis for each of the specimens was consistent with GBM.

Postoperative Treatment

The patient underwent adjuvant radiotherapy (56 Gy over 6 weeks) and adjuvant chemotherapy with temozolomide, which was initially administered concomitantly with radiotherapy (75 mg/m²/day) and then sequentially.
(150 mg/m²/day for 5 days in each cycle of 28 days, for 3 cycles). The patient subsequently underwent chemotherapy with the administration of temozolomide (75 mg/m²/day in each cycle of 21 days) along with bevacizumab (10 mg/kg once every 14 days) for 6 months. The patient showed clear signs of clinical deterioration at 10 months and succumbed to the disease at 12 months.

CGH Array and RNA Sequencing

To interrogate copy number aberrations, gene expression similarities and differences, and to explore a possible lineage among the four tumors, comparative genomic hybridization (CGH) array (aCGH) and transcriptome sequencing (RNA-sequencing) were performed. We used the circular binary segmentation (CBS) algorithm to calculate copy number changes in aCGH data. Reads from RNA-sequencing data were aligned to hg19 (Human Genome version 19), and the assembly and expression values were calculated as RPKM (reads per kilobase per million mapped reads) values using bioscope 1.3 software by Life Technologies as described. Hierarchical clustering of 840 genes was performed using MeV 4.9.0. The 840 genes were used to determine the molecular subtype of GBM. The expression profiles were normalized using standard score and then a pairwise average-linkage clustering method was used and visualized in MeV (Fig. 3). As shown, frontal, temporal, and occipital horn tumor samples (SN132_1, SN132_2, SN132_4, respectively) clustered adjacent. The hippocampal tumor (SN132_3) expression profile was distantly related and showed greater similarity
to tumors resected from different GBM patients. Furthermore, RNA-sequencing and aCGH expression data (Fig. 4) showed a shared set of aberrations among the hippocampal, frontal, temporal, and occipital horn lesions. All 4 tumors harbored a R175H mutation in the TP53 gene, had focal amplification on chromosome 6 accompanied by overexpression of the HDAC2 and MARCKS genes in that locus, deletion of the CDKN2A/CDKN2B locus, and loss of expression of the MTSS1 gene due to focal deletion on chromosome 8. However, each of these masses also contained unique aberrations that made them highly differentiated. The hippocampal tumor had MET amplification, the frontal tumor had focal chromosome 3 amplification, the occipital tumor had focal amplification on chromosome 9, and the temporal tumor showed focal amplification of the EGFR locus and presence of an EGFRvIII mutation. These data suggested that all of the tumors had a common origin while each had evolved a unique genetic signature. Thus, a diagnosis of multicentric GBM was made.

**Discussion**

Treating patients with multiple GBM lesions has been challenging for oncologists and surgeons, especially since no definitive treatment guidelines exist. The incidence of multiple and simultaneous brain cancer foci upon diagnosis has historically been reported as between 0.5% and 20%. However, a recent study found that 35% of patients had multiple lesions at the time of diagnosis. This prevalence necessitates further investigation.

Of note, the clinical strategy chosen in our case—resecting multiple lesions—is more aggressive than usual. The patient had 3 children and was in otherwise good health. Also, the resections were not performed for investigative purposes. It has been reported that the use of multiple craniotomies for multiple lesions is not associated with increased risks or complications (vs a single resection) and that the duration of survival for GBM patients with multiple lesions in which all lesions were resected was similar to that of patients in a matched cohort who underwent resection of a single glioblastoma.

Here we present a patient with 4 GBM tumors that were shown through in-depth genetic investigation to have a small degree of similarity and a marked level of genetic heterogeneity (when compared with one another). The data suggest that the 4 tumors share a common origin, yet each acquired a unique set of aberrations. Interestingly, even though 3 of the tumors were of the mesenchymal subtype, they were no more similar than tumors of the same subtype from different patients.

In an era of personalized medicine based on technological advancements, greater genetic interrogation is needed to produce more meaningful long-term survival data, particularly in patients with multiple glioblastoma lesions. Therapeutic advances have done little to minimize disease progression or to improve survival for this subpopulation of patients. In cases of multicentric and multifocal disease, genetic investigation of each tumor may permit more precise tumor characterizations, creating highly targeted therapies to improve patient outcomes.

**FIG. 3.** Pairwise average-linkage clustering of RNA-sequencing data. Hierarchical clustering of 840 gene expression profiles from RNA-sequencing data (28 GBM and 4 nontumor brain samples) was performed using MeV 4.9.0. The 840 genes were used to determine the molecular subtype of GBM. Frontal, temporal, and occipital horn tumor samples (SN132_1, SN132_2, SN132_4, respectively) clustered adjacently as mesenchymal subtype (blue bar). The hippocampal tumor (SN132_3) expression profile was distantly related, had proneural subtype (magenta bar), and showed greater similarity to tumors resected from different GBM patients with proneural subtype. Figure is available in color online only.
FIG. 4. Tumor lineage based on RNA-sequencing and aCGH data. All 4 tumors harbor an R175H mutation in the TP53 gene, have overexpression of the HDAC2 and MARCKS genes due to focal amplification on chromosome 6, deletion of the CDKN2A/CDKN2B locus, and loss of expression of several genes, including MTSS1, due to focal deletion on chromosome 8. The hippocampal tumor has MET overexpression and amplification. The frontal, occipital, and temporal lobe tumors share loss of expression of SLC1A1 due to chromosomal loss. The frontal lobe tumor has focal chromosome 3 amplification accompanied by overexpression of several genes in that locus. The occipital and temporal lobe tumors share deletion of the LRP1B, ZFYVE26, and RAD51L1 genes. The occipital lobe tumor has focal amplification on chromosome 9 accompanied by overexpression of several genes in that locus, and the temporal lobe tumor shows focal amplification of the EGFR locus and presence of EGFRVIII mutation. Amp = amplification; del = deletion; chr = chromosome. Figure is available in color online only.

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References
8. Louis DN, Ohgaki H, Wiestler OD, Cavenee WK, Burger PC,

Disclosure
The authors report no conflict of interest concerning the materials or methods used in this study or the findings specified in this paper.

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