Magnetic resonance spectroscopy detection of high lipid levels in intraaxial tumors without central necrosis: a characteristic of malignant lymphoma

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OBJECT The differentiation of malignant lymphomas from gliomas or malignant gliomas by conventional MRI can be difficult. The authors studied Gd-enhanced MR images to obtain a differential diagnosis between malignant lymphomas and gliomas without central necrosis or cystic changes and investigated the diagnostic value of single-voxel proton MR spectroscopy (1H-MRS) using different parameters, including lipid levels.

METHODS This was a retrospective study of patients with primary malignant CNS lymphoma (n = 17) and glioma (n = 122 [Grades I, II, III, and IV in 10, 30, 33, and 49 patients, respectively]) who were treated between 2007 and 2013. The authors focused on 15 patients with homogeneously enhanced primary malignant CNS lymphomas and 7 homogeneously enhanced gliomas. Images of all the included tumors were acquired with 1H-MRS at 3 T, and the diagnoses were histologically confirmed.

RESULTS Using a short echo time 1H-MRS, large lipid peaks were observed in all 17 patients with a malignant lymphoma, in 39 patients (79.6%) with a Grade IV glioma, and in 10 patients (30.3%) with a Grade III glioma. A focus on homogeneously enhanced tumors revealed large lipid peaks in 15 malignant lymphomas that were free of central necrosis on Gd-enhanced T1-weighted images. Conversely, in the 7 homogeneously enhanced gliomas (glioblastoma and anaplastic astrocytoma, n = 2 each; anaplastic oligodendroglioma, diffuse astrocytoma, and pilomyxoid astrocytoma, n = 1 each), lipid peaks were small or absent.

CONCLUSIONS Large lipid peaks on 1H-MRS images of tumors without central necrosis were characteristic of malignant lymphomas. Conversely, small or absent lipid peaks in intraaxial tumors without central necrosis were strongly suggestive of glioma.

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KEY WORDS malignant lymphoma; proton MR spectroscopy; lipids; glioma; oncology

Primary CNS lymphomas, a rare malignant form of non-Hodgkin’s lymphoma, account for 3%–6% of all primary brain neoplasms. There has been an increase in the incidence of malignant lymphoma in the last 3 decades. The differentiation of malignant lymphomas from gliomas or malignant gliomas by conventional MRI can be difficult. Malignant lymphomas and glioblastomas require different treatments; glioblastomas are treated by extensive resection, whereas malignant lymphomas are usually addressed by biopsy. Therefore, a differential diagnosis is of high clinical relevance. Moreover, even with advanced MRI techniques that yield physiological in-
formation, it is often difficult to differentiate malignant lymphomas from nonneoplastic diseases such as neuro-Behçet disease, multiple sclerosis, and acute demyelinating encephalomyelitis.

Magnetic resonance spectroscopy (MRS) is a noninvasive technique for obtaining biochemical information from the volume of interest (VOI) in tissues. In vivo proton MRS (1H-MRS) provides important information on tumor activity and tumor-tissue characteristics. The usefulness of 1H-MRS parameters in the differential diagnosis of brain tumors has been reported. Among these parameters, lipid peaks can be effectively detected, especially at a short echo time. In brain tumors, the presence of lipids often indicates the presence of necrotic tissue, which is considered to be an indicator of malignancy and a poor prognosis. Because malignant brain tumors tend to exhibit central necrosis on Gd-enhanced T1-weighted images, central necrosis inside the VOI is reflected by lipid peaks on 1H-MRS images. For the analysis of specific tumor characteristics, it is important to select the VOI only in Gd-enhanced lesions. However, because the signal-to-noise ratio from the pixel level of the VOI is low, it is difficult to avoid signals from necrotic or cystic lesions even when using the multivoxel 1H-MRS technique.

We used Gd-enhanced MRI to establish a differential diagnosis between malignant lymphomas and gliomas without central necrosis or cystic changes and examined the diagnostic value of 1H-MRS from the perspective of lipid peaks. We found that the presence on short echo time 1H-MRS images of large lipid peaks in tumors without central necrosis is a characteristic of malignant lymphomas.

Methods

Patients

This retrospective study was approved by the Hiroshima University Hospital’s institutional review board; the requirement for written patient consent was waived. To protect patient privacy, we removed all identifiers from our records after the completion of our analyses.

Between 2007 and 2013, we treated 30 patients with malignant lymphoma at our institution. We excluded 13 tumors because they were metastatic malignant lymphomas (n = 7) or their size was too small for setting a VOI (n = 6). Consequently, 17 malignant lymphomas (in 6 males and 11 females), 2 of which were tumors with central necrosis, were included in the study. The 15 lymphomas without central necroses were homogeneously enhanced on T1-weighted images; they were the focus of this study.

In that same period, we also treated 143 patients with newly diagnosed surgically confirmed glioma. We excluded 21 tumors because they were not examined by 1H-MRS (n = 12), their size was too small for setting a VOI (n = 3), or their spectra were of poor quality (n = 6), as indicated by an increased line width of the water resonance by 3 or more SDs above the mean line width of all tumor spectra. Consequently, our study included 122 gliomas (in 71 males and 51 females); 7 were homogeneously enhanced on T1-weighted images and were the focus of this study.

MRI Studies

All MRI studies were performed using a 3.0-T superconducting system (Signa Excite HD 3.0 T, versions 12 [January 2007 to March 2010] and 15 [April 2010 onward]; GE Medical Systems). The MRI studies were performed in 3 orthogonal planes and included nonenhanced T1-weighted images (TR 450 msec, TE 18 msec, FOV 22 × 22 cm, matrix size 256 × 192/1 excitation, section thickness 6 mm, intersection gap 1.0 mm, 2 acquisitions), T2-weighted images (TR 4800 msec, TE 100 msec, echo train length 18, FOV 22 × 22 cm, matrix size 512 × 320/2 excitations, section thickness 6 mm, intersection gap 1.0 mm, 1 acquisition), FLAIR images (TR 10,000 msec, TE 140.0 msec, inversion recovery time 2400.0 msec, FOV 22 × 22 cm, matrix size 288 × 160/1 excitations, section thickness 6 mm, intersection gap 1.0 mm, 2 acquisitions), and T2*-weighted images (TR 600.0 msec, TE 12 msec, FOV 22 × 22 cm, matrix size 320 × 192/1 excitation, section thickness 6 mm, intersection gap 1.0 mm, 1 acquisition).

Diffusion-weighted imaging (DWI) was performed with b values of 1000 and 4000 sec/mm² (b-1000 and b-4000, respectively). The effective gradient was 40 mT/m, and the slew rate was 150 mT/m/msec. We used an 8-channel phased-array head coil with the following imaging parameters: TR 5000 msec and TE 73.2 msec (b=1000), TR 5000 msec and TE 100 msec (b=4000), number of excitations 1, FOV 22 × 22 cm, slice thickness 6 mm, intersection gap 1.0 mm, number of slices 20, data acquisition matrix 128 × 128, and 2 acquisitions. The scan times were 20 and 40 seconds for b-1000 and b-4000, respectively.

Transverse, sagittal, and coronal spin-echo T1-weighted images were acquired after the intravenous administration of a Gd-based contrast medium and using the following parameters: TR 450 msec, TE 18 msec, FOV 18 × 18 cm, matrix size 288 × 192, section thickness 6 mm, intersection gap 1.0 mm, and 2 acquisitions. The Gd-enhanced pattern of each tumor was defined as having no, focal, heterogeneous, ring, or homogeneous enhancement.

Proton MRS

For 1H-MRS, we applied the point-resolved spectroscopic method (probe-P; TR 2000 msec, TE 144 msec, 2048 data points, 64 signals acquired) with a circularly polarized head coil. Additional spectra with a short (30-msec) TE were also acquired. To guide the single-voxel spectroscopic examinations, we used enhanced T1-weighted images for enhanced tumors and T2-weighted FLAIR images for nonenhanced tumors. Under 3D control, the rectangular 1H-MRS voxel was placed on the solid tumor area; efforts were made to avoid contamination by surrounding normal-appearing tissue, the skull base, any hemorrhagic lesions, and the ventricular system and to include enhanced tumor lesions in patients with an enhanced tumor and FLAIR high-intensity lesions in patients with nonenhanced tumor. The voxel size was between 15 × 15 × 15 mm (volume 3.4 cm³) and 20 × 20 × 20 mm (volume 8 cm³); the estimated disease fraction in the VOI exceeded 60%–100% in each patient. Spatial suppression pulses were applied to the outside of the voxel to reduce spectral contamination; global and localized shimming on the

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water proton and optimization of water suppression were performed, resulting in water-peak line widths of 2–4 Hz. All MR images covering the VOI selected for MRS were retrospectively assessed consensually by 3 of the authors (F.Y., T.T., and R.N.). Spectra of poor quality, identified by an increased line width of the water resonance (a measure of the field homogeneity in the VOI) of 3 or more SDs above the mean line width of all tumor spectra, were excluded from the final analyses. The line width of the water signal intensity was automatically measured and reported by the processing software. The relative signal intensities of choline (Cho), creatine (Cr), N-acetyl aspartate (NAA), and myoinositol (mI) were obtained by numeric integration of the fitted signals. The signals of lipid peaks were determined by their short TEs, because the relaxation time of a lipid is very short. Lipid peaks were defined as negative and undetectable (–), positive and detectable (1+), strongly positive (2+, representing the second largest among all the peaks), and extremely strongly positive (3+, representing the largest among all the peaks).

### Apparent Diffusion Coefficient Maps and Calculations

All apparent diffusion coefficient (ADC) maps were generated with software (FuncTool; GE Medical Systems). They were obtained by calculating the signal intensities on diffusion-weighted images at 2 different b values (0 and 1000 sec/mm² for b-1000 ADC maps and 0 and 4000 sec/mm² for b-4000 ADC maps) on a pixel-by-pixel basis. To determine the ADC, 2 independent readers (A.D. and F.Y.) manually placed regions of interest (ROIs) in tumor regions on b-1000 ADC maps and as close as possible in the same region of an approximately similar size (approximately 30 mm²) on the b-4000 ADC maps. The ROI were placed on enhanced lesions on contrast-enhanced T1-weighted MR images, as central as possible within the tumor. The number of ROI placed depended on the size of the observable part of the enhanced lesion. The ADC values of each tumor, based on 3–10 ROIs on the b-1000 and b-4000 ADC maps, were calculated, and the minimum absolute ADC (ADCMIN) values were obtained.

### Histological Study

Tumor specimens obtained by resection or biopsy were fixed in 10% phosphate-buffered formalin and embedded in paraffin. Representative slides were stained with H & E for a standard histological diagnosis according to WHO criteria by the consensus of 2 of the authors (V.J.A. and Y.T.), who were blinded to all clinical and radiological data.
Statistical Analysis

Statistical analyses were performed with StatView software (version 5.0; SAS Institute). The relationship between MRS parameters that included the NAA-to-Cr, Cho-to-Cr, Cho-to-NAA, and mI-to-Cr ratios, the lipid levels, the averaged ADCs, and the different brain tumor types was evaluated with logistic analysis, the chi-square test, and the Mann-Whitney U-test; statistical significance was assigned when the p value was < 0.05.

Results

Patients and Glioma Grades

The patients with a malignant lymphoma or glioma ranged in age from 36 to 78 years (mean 63.9 years, median 66 years) and from 1 to 85 years (mean 46.5 years, median 48.5 years), respectively. Of the 122 gliomas, 49 were Grade IV (47 glioblastomas and 2 gliosarcomas), 33 were Grade III (12 anaplastic astrocytomas, 12 anaplastic oligoastrocytomas, 8 anaplastic oligodendrogliomas, 1 anaplastic pleomorphic xanthoastrocytoma), 30 were Grade II (5 diffuse astrocytomas, 13 oligoastrocytomas, 8 oligodendrogliomas, 1 anaplastic pleomorphic xanthoastrocytoma), 7 were Grade I (2 pilocytic astrocytomas, 1 ganglioglioma, and 4 pilomyxoid astrocytomas).

Gadolinium enhancement was absent in 21, focal in 21, heterogeneous in 25, and homogeneous in 7 of the 122 gliomas; 47 were ring enhanced. For 1 patient with a glioblastoma and renal failure, we did not attempt to obtain enhanced images; the VOI was chosen on T2-weighted FLAIR images.

Lipid Peaks Detected by Single-Voxel 1H-MRS

We first analyzed the lipid peaks of gliomas and lymphomas seen in 1H-MRS images (Table 1). In each of the 17 malignant lymphomas, we observed strong-positive (2+) or extremely strong-positive (3+) lipid peaks. Of the 122 Grade I–IV gliomas, 52 (42.6%) manifested 2+ or 3+ lipid peaks.

Although lipid peaks were rare in lower-grade gliomas, we observed 2+ and 3+ peaks in 30.3% of the Grade III and 79.6% of the Grade IV gliomas (low- vs high-grade gliomas, p < 0.0001, chi-square test; Grade III vs IV, p < 0.0001, chi-square test). No unenhanced gliomas showed lipid peaks. The expression of 2+ or 3+ lipid peaks was statistically higher in malignant lymphomas than in high-grade gliomas (p = 0.0014, chi-square test).

1H-MRS Findings in Ring-Enhanced Tumors

Next, we focused on ring-enhanced tumors (Table 2). Both of the ring-enhanced malignant lymphomas showed 3+ lipid peaks. We observed 2+ or 3+ lipid peaks in 43 of the 46 ring-enhanced gliomas. All but one glioma, a ring-enhanced pilocytic astrocytoma, showed lipid peaks.

1H-MRS Findings of Homogeneously Enhanced Tumors

We then focused on the patients with homogeneously enhanced lymphomas and gliomas. Our MRS findings on malignant lymphomas and gliomas without central necrosis are summarized in Table 3. All of the malignant lymphomas but none of the gliomas showed 2+ or 3+ lipid expression (Figs. 1 and 2) (p = 0.003, chi-square test). Representative gliomas are shown in Figs. 3–5. When we analyzed the diagnostic value of 1H-MRS with respect to other parameters, including Cho, Cr, NAA, and mI, we detected no difference between malignant lymphomas and gliomas. Logistic analysis revealed that lipid expression differentiated between malignant lymphomas and gliomas and that logistic discriminant analysis clearly separated these tumors.

### TABLE 2. Lipid expression levels of the ring-enhanced gliomas and malignant lymphomas

<table>
<thead>
<tr>
<th>Tumor Type &amp; Grade</th>
<th>No. of Patients</th>
<th>No. (%) w/ Lipid Level of:</th>
<th>1+</th>
<th>2+</th>
<th>3+</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ring-enhanced gliomas (n = 46)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Grade I</td>
<td>1</td>
<td>1 (100)</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Pilocytic astrocytoma</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Grade II, total</td>
<td>0</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Grade III</td>
<td>7</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>4 (57.1)</td>
<td>3 (42.9)</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anaplastic astrocytoma</td>
<td>2</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Anaplastic oligoastrocytoma</td>
<td>2</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Anaplastic oligodendroglioma</td>
<td>2</td>
<td>0</td>
<td>0</td>
<td>2</td>
<td>0</td>
</tr>
<tr>
<td>Anaplastic PXA</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
</tr>
<tr>
<td>Grade IV</td>
<td>38</td>
<td>0 (0)</td>
<td>2 (5.3)</td>
<td>1 (2.6)</td>
<td>35 (92.1)</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Glioblastoma</td>
<td>38</td>
<td>0</td>
<td>2</td>
<td>1</td>
<td>35</td>
</tr>
<tr>
<td>Ring-enhanced lymphomas (n = 2)</td>
<td>2</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>2 (100)</td>
</tr>
</tbody>
</table>
Finally, we checked the diagnostic value of DWI and the ADCs of homogeneously enhanced tumors. The ADC results for malignant lymphomas and gliomas without central necrosis are summarized in Table 3. Comparisons at b-1000 and b-4000 showed that the ADC MIN values of gliomas were higher than those of malignant lymphomas (p = 0.0220 and 0.0010, respectively, Mann-Whitney U-test) (Fig. 6). However, there was some overlap between gliomas and malignant lymphomas at both b values.

### Discussion

Our results show that high lipid peaks in tumors without central necrosis are a characteristic of malignant lymphomas and that their absence in intraaxial tumors without central necrosis is strongly suggestive of glioma. Although others reported that malignant lymphomas demonstrated massively elevated lipid resonances, some patients with high-grade gliomas, including glioblastomas, manifested high lipid peaks.3,12,16,25,26 Our findings on Grade I–IV gliomas, especially ring-enhanced glioblastomas, are consistent with theirs. The earlier studies did not focus on homogeneously enhanced lesions; rather, they analyzed heterogeneously enhanced, ring-enhanced, and nonenhanced tumors together. Although it is important to set the ROI on enhanced tumor lesions and to avoid placement on nonenhanced lesions (e.g., areas of central necrosis), it can be difficult. Therefore, we focused on tumors without central necrosis (i.e., homogeneously enhanced tumors). We found that there was a characteristic difference in the presence and size of lipid resonance peaks on 1H-MRS images between homogeneously enhanced gliomas and malignant lymphomas. Consequently, although our and earlier studies showed a statistically significant difference in lipid expression between malignant lymphomas and high-grade gliomas, our findings on homogeneously enhanced tumors show that lipid expression differentiates between malignant lymphomas and high-grade gliomas. This finding is clinically relevant because homogeneously enhanced gliomas can be differentiated from malignant lymphomas by the absence of high lipid resonance peaks on 1H-MRS images.

### Table 3: Malignant lymphomas and gliomas without central necrosis

<table>
<thead>
<tr>
<th>Tumor Type</th>
<th>Age (yrs), Sex</th>
<th>ADC Min From DWI</th>
<th>ADC Min</th>
<th>VolI-Included Tumor Vol (%)</th>
<th>TE 144 msec</th>
<th>TE 30 msec</th>
</tr>
</thead>
<tbody>
<tr>
<td>Homogeneously enhanced intraaxial tumors</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>62, F</td>
<td>0.854</td>
<td>0.462</td>
<td>60</td>
<td>26, 21, 24</td>
<td>ND 55, 47</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>76, F</td>
<td>0.592</td>
<td>0.352</td>
<td>60</td>
<td>32, 15, 14</td>
<td>ND 34, 40</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>63, F</td>
<td>0.648</td>
<td>0.420</td>
<td>60</td>
<td>29, 15, 24</td>
<td>ND 79, 42</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>64, M</td>
<td>0.558</td>
<td>0.346</td>
<td>60</td>
<td>21, 8, 10</td>
<td>ND 39, 24</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>72, F</td>
<td>0.795</td>
<td>0.463</td>
<td>60</td>
<td>27, 8, 12</td>
<td>ND 74, ND</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>58, F</td>
<td>0.835</td>
<td>0.469</td>
<td>60</td>
<td>41, 12, 14</td>
<td>ND 97, 35</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>39, F</td>
<td>0.645</td>
<td>0.385</td>
<td>80</td>
<td>47, 9, 13</td>
<td>ND 93, 27</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>68, M</td>
<td>0.593</td>
<td>0.386</td>
<td>80</td>
<td>59, ND, ND</td>
<td>ND 113, ND</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>60, M</td>
<td>0.499</td>
<td>0.260</td>
<td>80</td>
<td>32, 7, ND</td>
<td>ND 78, ND</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>72, F</td>
<td>0.693</td>
<td>0.389</td>
<td>80</td>
<td>76, 23, 25</td>
<td>153, 56, 86</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>67, F</td>
<td>0.726</td>
<td>0.425</td>
<td>80</td>
<td>27, ND, ND</td>
<td>ND 76, 20</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>63, F</td>
<td>0.863</td>
<td>0.523</td>
<td>80</td>
<td>20, ND, ND</td>
<td>ND 75, ND</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>76, F</td>
<td>0.626</td>
<td>0.366</td>
<td>80</td>
<td>44, ND, ND</td>
<td>ND 82, ND</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>66, M</td>
<td>0.668</td>
<td>0.322</td>
<td>80</td>
<td>63, ND, ND</td>
<td>ND 113, ND</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>78, F</td>
<td>0.542</td>
<td>0.350</td>
<td>80</td>
<td>31, 9, ND</td>
<td>68, 43, ND</td>
</tr>
<tr>
<td>Gliomas</td>
<td></td>
<td></td>
<td></td>
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<td></td>
<td></td>
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<tr>
<td>Diffuse astrocytoma</td>
<td>47, M</td>
<td>1.10</td>
<td>0.594</td>
<td>60</td>
<td>54, 9, 14</td>
<td>ND 99, 19</td>
</tr>
<tr>
<td>Anaplastic oligodendroglioma</td>
<td>48, F</td>
<td>0.786</td>
<td>0.481</td>
<td>60</td>
<td>88, 30, 46</td>
<td>ND 152, 71</td>
</tr>
<tr>
<td>Pilomyxoid astrocytoma</td>
<td>1, F</td>
<td>1.92</td>
<td>1.09</td>
<td>80</td>
<td>43, 10, 28</td>
<td>ND 71, 29</td>
</tr>
<tr>
<td>Anaplastic astrocytoma</td>
<td>54, F</td>
<td>0.704</td>
<td>0.530</td>
<td>80</td>
<td>46, 9, 14</td>
<td>77, 34, 37</td>
</tr>
<tr>
<td>Anaplastic astrocytoma</td>
<td>65, F</td>
<td>1.30</td>
<td>0.795</td>
<td>80</td>
<td>44, 12, ND</td>
<td>85, 33, 45</td>
</tr>
<tr>
<td>Glioblastoma</td>
<td>31, M</td>
<td>0.705</td>
<td>0.495</td>
<td>80</td>
<td>128, 16, 13</td>
<td>199, ND, ND</td>
</tr>
<tr>
<td>Glioblastoma</td>
<td>66, F</td>
<td>0.690</td>
<td>0.457</td>
<td>80</td>
<td>92, 51, 38</td>
<td>183, 115, 119</td>
</tr>
</tbody>
</table>

ND = not detected.
The detection of high lipid peaks in malignant lymphomas has been suggested to reflect a high cell turnover and infiltration by macrophages. The high lipid levels in these tumors without necrosis may be caused by the presence of activated or transformed lymphocytes or macrophages, because these cell types can contain high levels of MR-visible lipids. Primary CNS lymphomas are hypercellular and comprise lymphoid cells and macrophages. Phagocytosis of cell membranes by infiltrated macrophages results in lipid signals. Lipid signals have...
also been attributed to in vivo apoptosis or programmed cell death. Fluorescence microscopy and flow-cytometry measurements have shown that an increase in 1H-MRS-detectable lipids was correlated with an accumulation of cytoplasmic lipid droplets, the so-called lipid bodies. Lipid signals in most brain tumors can be observed when the spectra are acquired at a TE of < 50 msec. Mobile lipid resonances are generally observed in contrast-enhancing and necrotic areas of the tumor. Brain tumors tend to be highly heterogeneous and include areas of viable tumor, necrosis, and hemorrhage. Elevated lipid peaks are a characteristic of solid and cystic high-grade gliomas but not of

**Fig. 3.** Glioblastoma in a 31-year-old man. A: Axial contrast-enhanced T1-weighted image showing diffuse tumor enhancement in the thalamus bilaterally. B: Axial FLAIR image showing tumor isointensity to high intensity and hyperintense peritumoral edema. C: Single-voxel 1H-MRS image (TE 35 msec) showing a tumor with a positive lipid peak (+). D and E: Axial diffusion-weighted images at b-1000 and b-4000, respectively, showing tumor hyperintensity. F: Photomicrograph of H & E–stained tumor tissue. Original magnification ×400. Figure is available in color online only.

**Fig. 4.** Glioblastoma in a 66-year-old woman. A: Axial contrast-enhanced T1-weighted image showing diffuse tumor enhancement in the right parietal white matter and a small enhanced lesion in the bilateral parietal lobe. B: Axial FLAIR image showing tumor high intensity and hyperintense deep white matter. C: Single-voxel 1H-MRS image (TE 35 msec) showing a tumor negative for lipid peaks. D and E: Axial diffusion-weighted images at b-1000 and b-4000, respectively, showing tumor hyperintensity. F: Photomicrograph of H & E–stained tumor tissue (magnification ×400). Figure is available in color online only.
High lipid levels in malignant lymphoma on \textsuperscript{1}H-MRS

Among our homogeneously enhanced gliomas, low- and high-grade tumors did not differ with respect to lipid peaks, which suggests that the higher lipid peaks in high-grade gliomas may be attributable to their cystic or necrotic components. Future studies are necessary to assess the relationship between the size of lipid peaks and glioma grade.

The minimum or mean ADCs of malignant lymphomas are statistically lower than those of gliomas.\textsuperscript{6,10,15,20,32,35} The sensitivity and specificity of the ADC for discriminating between malignant lymphomas and gliomas are approximately 80\%–90\%. However, some studies reported no statistically significant difference in the ADCs of glioblastomas and malignant lymphomas, although the ADCs of malignant lymphomas were lower than those of high-grade gliomas.\textsuperscript{24,31} In fact, the overlap in ADC values rendered the differentiation between malignant lymphomas and gliomas on the basis of the ADC difficult.\textsuperscript{20,31,35} For better and more accurate differentiation, other methods (e.g., fractional anisotropy,\textsuperscript{22} perfusion-weighted imaging,\textsuperscript{37} MRS, and PET imaging\textsuperscript{24,37}) were added to the ADC parameters. Although they were useful, some overlapping between the malignant lymphomas and gliomas persisted.

A differential diagnostic advantage of higher b-value-based DWI in many CNS diseases, including malignant lymphomas, has been reported.\textsuperscript{6} Although we found that b-4000\--based DWI was superior to b-1000\--based DWI, it was not possible to differentiate all malignant lymphomas from gliomas even with the higher b value.

Our study has some limitations. We focused strictly...
on the differentiation between malignant lymphomas and gliomas. We did not include tumefactive demyelinating lesions or metastatic tumors in our analysis because, at our institution, all metastatic tumors larger than 2 cm contained some cystic formations or central necrosis. We could not apply our single-voxel ¹H-MRS method to study small tumors. Future studies are necessary to confirm the lipid expression of homogeneously enhanced metastases. Although here we used qualitative rather than quantitative methods to evaluate lipid signals, quantitative studies using LCModel software are in progress in our laboratory.

Conclusions

¹H-MRS is useful for differentiating between gliomas and malignant lymphomas. The ¹H-MRS detection of large lipid peaks in tumors without central necrosis is a characteristic of malignant lymphomas. The detection of no or small lipid peaks in an intraaxial tumor without central necrosis is strongly suggestive of glioma.

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References

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