Certain types of tumors, in the CNS and elsewhere, secrete substances that can be measured in the peripheral blood, and these biological markers routinely serve to screen, to diagnose, or to follow the course of the underlying disease. So far it has not been established if the fraction of tumor removal can be accurately determined by peripheral blood sampling. If there existed a close correlation between tumor mass and the serum level of a material secreted by the tumor, and if this substance could be easily and reliably measured in the peripheral blood, it would permit easy calculation of the remaining tumor mass after surgical removal or other ablative therapies.

Acromegaly is associated with considerable morbidity and mortality. First-line management is transsphenoidal tumor resection with the goal to normalize the growth hormone (GH) and insulin-like growth factor–1 (IGF-1) levels. However, since many of these tumors invade surrounding structures total resection cannot always be achieved. Hence, the hormone levels in these cases decrease after partial tumor removal, but do not become normal.

Although GH-secreting tumors produce levels of GH that can be easily measured in the peripheral blood, the correlation between tumor size and plasma GH levels varies widely from patient to patient. Furthermore, although there is a relationship between GH and IGF-1, it is nonlinear because of saturation of GH receptors when GH exceeds certain threshold levels, thresholds that also seem to vary widely from patient to patient.

Abbreviations

GH = growth hormone; IGF-1 = insulin-like growth factor–1.

Submitted March 4, 2014. Accepted October 9, 2014.

Include When Citing Published online November 28, 2014; DOI: 10.3171/2014.10.JNS14496.

Disclosure Dr. Schwyzer was supported by a postdoctoral fellowship grant from the Emil-Boral Foundation of Austria and Switzerland.
To investigate this further, we examined various relationships between tumor size, plasma GH levels, and plasma IGF-1 before and after transsphenoidal surgery in patients with acromegaly caused by GH-secreting tumors.

We explored the possibility that if individual tumors have their own intrinsic level of GH production and if that level of GH production is homogeneous across the tumor, a comparison of GH levels before and after surgery would indicate the fraction of tumor that had been removed. Thus, a close correlation between tumor volume and hormone secretion in individual patients would permit calculation of the fraction of tumor removed by surgery, simply by measuring the postoperative GH levels.

**Methods**

We assessed the GH and IGF-1 levels before and after surgery in patients who were not on medical therapy and who had incomplete tumor removal, permitting the measurement of tumor mass before and after surgery. With approval from our local institutional review board, prospectively recorded medical records and imaging studies were systematically reviewed. Patients were excluded 1) if they had no visible residual tumor on postoperative MRI, because there could be no measurement of tumor size after surgery; and 2) if they were defined by the contemporary endocrine criteria as being in remission after surgery (IGF-1 in the normal range for age and sex, glucose-suppressed plasma GH < 1.0 ng/ml), because the pulsatile nature of GH secretion after surgical remission makes random GH measurement unreliable.6 Patients underwent surgery between 2006 and 2009. The 3D region of interest–based volumetric method, in which the tumor margin is traced on each MRI slice and then the volume is calculated by using the area of each tumor on each slice and the slice thickness, was used to measure the tumor volume before and after surgery (NIH ImageJ (http://rsbweb.nih.gov/ij/)).

The values of GH and IGF-1 used were the last measurements before surgery (for GH 1–4 days before surgery, except in 1 patient it was 1 month before surgery; for IGF-1, 1–3 days before surgery, except in 2 patients it was 1 month before surgery); postoperative GH was obtained at 6–7:00 in the morning of the 1st day after surgery in all patients, except in 1 patient it was at 1 month, because no immediate postoperative GH was obtained. Because of its prolonged half-life of several weeks in the peripheral blood, postoperative IGF-1 was measured 2–3 months after surgery in all but 1 patient, in whom it was obtained 6 months after surgery. Tumor volumes were obtained on the last MRI before surgery (1 day–2 months before surgery) and on the first MRI obtained after surgery (2–3 months after surgery). Two patients did not have IGF-1 measurements while off medical therapy at 2–3 months after surgery. The GH and IGF-1 assays were used and after surgery, permitting normalization of hormone levels per mass of tumor for each patient.

**Statistical Analysis**

Comparison of tumor subtype with peripheral GH and IGF-1 levels was done with the Kruskal-Wallis test, and correlations of GH, IGF-1, tumor volume, and fraction of tumor removed versus fraction of change in GH and IGF-1 were analyzed with the Spearman rank correlation test (rₚ) using GraphPad Prism (GraphPad Software).

Measurement of the tumor volumes was done by L.S., who performed the measurements blinded to the GH or IGF-1 levels.

**Results**

There were 11 patients who met the criteria for study among 96 patients who had surgery for acromegaly between 2006 and 2009.

Although there was a trend toward correlation between tumor size and GH, it was not significant (p = 0.07); there was a correlation between tumor volume and plasma IGF-1 (rₚ = 0.64, p = 0.002; Fig. 1). There was also a correlation when comparing plasma GH versus plasma IGF-1 (rₚ = 0.66, p = 0.001; Fig. 2). Comparison of percent tumor resection and percent drop in IGF-1 did not demonstrate a correlation (p = 0.42; Fig. 3). Despite that there was no significant correlation between tumor size and GH levels, comparison of percent tumor resection with percent drop in plasma GH, thus normalizing each tumor to its own initial volume and plasma GH level, yielded a high correlation (rₚ = 0.78, p = 0.006; Fig. 3). Densely granulated tumors were associated with greater values for peripheral GH levels in comparison with sparsely granulated tumors (Kruskal-Wallis, p = 0.04); there was a trend for the same with IGF-1 levels, but it did not reach statistical significance (p = 0.3; Fig. 4).

**Discussion**

Because plasma IGF-1 has a long half-life it has a relatively steady level in normal subjects and in patients with acromegaly. Thus, for diagnosing acromegaly, measurement of IGF-1 is more reliable than measurement of GH because of the random pulsatile nature of GH secretion. In acromegaly the pituitary tumor produces GH, whereas IGF-1 is produced in the liver in response to plasma GH; IGF-1 has a sigmoid-shaped relationship with GH as a result of the hormone-receptor interactions between GH and IGF-1, with IGF-1 reaching a plateau after GH levels have reached levels sufficient to saturate GH receptors. Studies in which frequent sampling of GH is performed indicate that although GH is secreted in pulses in patients with acromegaly, as it is in normal subjects, the secretory bursts in acromegaly are much less prominent in comparison with the elevated basal GH concentration.5,7,9 Thus, most random measurements of GH in patients with acromegaly reflect the relatively steady basal GH level, rather than a level associated with a secretory pulse. Since GH is produced by the tumor it seems logical that the plasma GH...
levels would correlate with the tumor size in individual patients, although if different tumors have different potency of GH production the slope of that correlation would differ from patient to patient.

Based on the cellular distribution of GH staining with immunohistochemistry and their appearance with electron microscopy, GH-producing tumors are classified as densely granulated, sparsely granulated, or as an intermediate, mixed group. Assessment of our small set of tumors indicates that the densely granulated tumors produce higher levels of plasma GH per unit of tissue volume than do sparsely granulated tumors. A similar trend occurred with plasma IGF-1 levels per unit of tumor volume. This is in agreement with the recent findings of Fougner et al., who in their recent analysis of 78 tumors concluded that densely granulated tumors produce more IGF-1 per unit of tumor volume than do sparsely granulated tumors. They noted a similar trend for GH, although it did not reach significance. Obari et al. had previously noted higher mean plasma GH levels in patients with densely granulated and mixed tumors compared with patients with sparsely granulated tumors, and Bakhtiar et al. had previously found that the GH-producing index (peripheral GH levels per cubic centimeter of tumor) was lower in sparsely granulated compared with densely granulated tumors.

That the percent reduction in GH levels after surgery is tightly linked to the percent of tumor that was resected has potential practical relevance. For instance, the success of radiation therapy and medical therapy for GH-producing tumors is related to the degree of pretreatment elevation of GH. Occasionally, preoperative imaging suggests that a tumor cannot be totally removed with surgery, but predicts that the fraction of tumor that can probably be safely removed would produce GH levels that would yield higher chances of achieving a normal GH and IGF-1 level with subsequent medical or irradiation therapy, and would permit earlier return to normal levels of GH and IGF-1 after adjuvant therapy. Knowing that the percent reduction in GH can be reasonably predicted by the fraction of tumor that can probably be removed surgically might provide information on whether to suggest additional surgery, or not.

The limitation of our study is the relatively small number of patients. However, to address the issue that we wished to address required that patient selection include

FIG. 1. Graphs showing a trend toward a correlation when comparing tumor volume to plasma GH (left; \( p = 0.07 \)). The levels of plasma IGF-1 correlated with tumor volume (right; \( r_s = 0.64, p = 0.002 \)). Note that the measurements include the volumes measured before and after surgery. The black closed circle in Figs. 1–3 represents the GH level of a patient whose GH was measured 1 month after surgery. All other postoperative GH measurements were made in the morning of the 1st day after surgery.

FIG. 2. Graph showing a correlation between plasma GH versus plasma IGF-1 (\( r_s = 0.66, p = 0.001 \)). The values plotted include the measurements before and after surgery. One GH value of 568 ng/ml is not included in the graph.
Fig. 3. Graphs showing that there was no correlation between the percent drop in plasma IGF-1 and the percent tumor removal (left; $p = 0.42$). However, a comparison of percent tumor resection with percent drop in plasma GH, permitting normalization of each tumor to its own plasma GH level, yielded a high correlation coefficient (right; $r_s = 0.78, p = 0.006$).

Fig. 4. Bar graphs showing that densely granulated tumors ($n = 4$) produced higher levels of plasma GH (left; Kruskal-Wallis, $p = 0.04$) and IGF-1 than sparsely granulated tumors ($n = 6$), although the difference did not reach statistical significance for IGF-1 (right; $p = 0.3$). The values shown represent the means of the average GH and IGF-1 levels per cc of tumor as measured before and after surgery. There was 1 mixed tumor. The error bars represent SEMs.
only those with partial tumor removal that could be imaged using MRI, in patients who were not receiving medical therapy during imaging performed before and after surgery.

Conclusions

These results indicate that each GH-secreting tumor has its own intrinsic level of GH production per mass of tumor, that the level is homogeneous over the tumor mass, and that this varies greatly between tumors. Our study further indicates that the fraction of a GH-secreting tumor incompletely removed by surgery can be accurately estimated simply by comparing plasma GH levels after surgery to those before surgery. The same principles will likely apply to other types of tumors that produce hormones or other products that can be measured in the peripheral blood, although we have been unable to find a similar analysis in any other tumor type.

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