Maternal administration of meclozine for the treatment of foramen magnum stenosis in transgenic mice with achondroplasia

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OBJECTIVE Achondroplasia (ACH) is the most common short-limbed skeletal dysplasia caused by gain-of-function mutations in the fibroblast growth factor receptor 3 (FGFR3) gene. Foramen magnum stenosis (FMS) is one of the serious neurological complications in ACH. Through comprehensive drug screening, the authors identified that meclozine, an over-the-counter drug for motion sickness, inhibited activation of FGFR3 signaling. Oral administration of meclozine to the growing ACH mice promoted longitudinal bone growth, but it did not prevent FMS. In the current study, the authors evaluated the effects of maternal administration of meclozine on FMS in ACH mice.

METHODS The area of the foramen magnum was measured in 17-day-old Fgfr3<sup>ach</sup> mice and wild-type mice using micro-CT scanning. Meclozine was administered to the pregnant mice carrying Fgfr3<sup>ach</sup> offspring from embryonic Day (ED) 14.5 to postnatal Day (PD) 4.5. Spheno-occipital and anterior intracerephal synchondroses were histologically examined, and the bony bridges were scored on PD 4.5. In wild-type mice, tissue concentrations of meclozine in ED 17.5 fetuses and PD 6.5 pups were investigated.

RESULTS The area of the foramen magnum was significantly smaller in 17-day-old Fgfr3<sup>ach</sup> mice than in wild-type mice (p < 0.005). There were no bony bridges in the sphenoid-occipital and anterior intracerephal synchondroses in wild-type mice, while some of the synchondroses prematurely closed in untreated Fgfr3<sup>ach</sup> mice at PD 4.5. The average bony bridge score in the cranial base was 7.05 ± 1.39 in untreated Fgfr3<sup>ach</sup> mice and 6.13 ± 2.03 in meclozine-treated Fgfr3<sup>ach</sup> mice. The scores were not statistically significant between mice with and those without meclozine treatment (p = 0.12). The average tissue concentration of meclozine was significantly higher (508.8 ± 205.2 ng/g) in PD 6.5 mice than in ED 17.5 mice (56.9 ± 20.05 ng/g) (p < 0.005).

CONCLUSIONS Maternal administration of meclozine postponed premature closure of synchondroses in some Fgfr3<sup>ach</sup> mice, but the effect on preventing bony bridge formation was not significant, probably due to low placental transmission of the drug. Meclozine is likely to exhibit a marginal effect on premature closure of synchondroses at the cranial base in ACH.

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ACHONDROPLASIA (ACH) is one of the most common skeletal dysplasias with disproportionate short stature caused by activating mutations in FGFR3 encoding the fibroblast growth factor receptor 3.10,11 In addition to rhizomelic shortening of the extremities, foramen magnum stenosis (FMS) is one of the most serious neurological complications in patients with ACH, which sometimes results in sudden unexpected death in infants.9 Cervicomedullary decompression surgery has been indicated for infants who exhibited lower-limb hyperreflexia or clonus on examination, central hypopnea demonstrated by polysomnography, and foramen magnum measures below the mean for children with ACH.8 Premature closure of sphenoid-occipital bone synchondrosis was confirmed by CT studies in patients with ACH.1 Partial closure of the sphenoid-occipital synchondrosis and anterior intracerephal...
synchondroses was also observed in 36-week-gestation fetuses with thanatophoric dysplasia,5 which represents the most severe phenotype among FGFR3 disorders. Experimentally, Fgfr3G374R+/mice corresponding to human ACH exhibited premature closure of synchondroses at the cranial base on PD 10, while there was no evidence of fused synchondroses in wild-type littermate mice.5 Therefore, growth-promoting treatment for FMS should be initiated before closure of occipital synchondroses. Several FGFR3-targeted treatments for ACH have been reported in recent years. Continuous intravenous administration of C-type natriuretic peptide (CNP) and CNP analog (BMN-111) was associated with significant bone growth recovery in a mouse model of ACH.2,15 Using induced pluripotent stem cells established from patients with FGFR3 disorders, Yamashita et al. demonstrated that statins also rescued the dwarf phenotype in an ACH model in mice.14 Through comprehensive drug screening, we identified that meclozine, an antihistamine drug that has been used as an anti–motion sickness medication for more than 50 years, inhibited FGFR3 signaling in various chondrocytic cell lines.4 In addition, meclozine increased longitudinal bone growth in an ACH mouse model by suppressing FGFR3 signaling. The effects of these FGFR3 inhibitors on FMS have not been studied in detail probably because of their postnatal administration.

For the treatment of FMS in ACH, we investigated the effects of maternally administered meclozine on premature closure of synchondroses in the cranial base in ACH mice.

**Methods**

**Mice**

Fgfr3ach mice (FVB/NJcl background) were provided by Dr. David M. Ornitz at Washington University in St. Louis.7 In brief, Fgfr3ach mice express activated FGFR3 in the growth plate using the Col2a1 promoter. In all experiments, we used transgenic mice carrying the heterozygous Fgfr3ach transgene. Due to the unavailability of a sufficient number of wild-type FVB/NJcl mice, we used C57BL/6 J wild-type mice to investigate the tissue concentration of meclozine. All experimental procedures were approved by the Animal Care and Use Committee at our institution.

**Foramen Magnum Measurements**

Seventeen-day-old wild-type mice and mutant mice were subjected to micro-CT scanning (0.5-mm Al filter, 50 kV, 500 μA for 0.054 seconds; SkyScan 1176, Bruker). Three-dimensional images from the CT scan were reconstructed by an in-house volume-rendering software.6 This software enabled us to render 3D views of the CT scan from arbitrary viewpoints and view directions as well as to measure the distance between 2 specific points. The areas of the foramen magnum were measured on reconstructed 3D images.

**Maternal Administration of Meclozine**

We previously demonstrated that the average plasma concentration of meclozine was 36.58 ± 20.12 ng/ml (± SD, n = 10) after intake of a 0.4 g/kg diet,5 which was below the mean peak drug concentration of 68.42 ng/ml after a single dose of 25 mg meclozine tablet in humans.13 According to our previous study, food containing meclozine was prepared by mixing 0.4 g of meclozine with 1 kg of food (Oriental Yeast Co.). Wild-type female mice were mated with Fgfr3ach male mice to produce normal and heterozygous mutant embryos. The pregnant mice were then randomly treated with meclozine from embryonic Day (ED) 14.5 to postnatal Day (PD) 4.5. The offspring were divided into 4 groups for each kinship: 1) meclozine-treated Fgfr3ach mice, 2) meclozine-treated wild-type mice, 3) untreated Fgfr3ach mice, and 4) untreated wild-type mice (Fig. 1).

**Skeletal Preparation**

Whole skeletons were harvested on PD 4.5 and stored in 90% ethanol for 3 days, followed by acetone treatment for 2 days.12 Specimens were then stained using Alizarin red to analyze ossified bones and Alcian blue to analyze

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**Fig. 1.** Flowchart showing the number of Fgfr3ach mice treated with or without meclozine. Pregnant wild-type mice mated with male Fgfr3ach mice were divided into 2 groups according to drug intervention. As a result, 16 meclozine-treated and 19 untreated Fgfr3ach mice were used.
cartilage for 3 days at 37°C. Following incubation, the samples were transferred to 1% KOH and incubated at room temperature for 2 days. The specimens were serially washed with decreasing concentrations (1% to 0%) of KOH and increasing concentrations (0% to 100%) of glycerol by monitoring the intensity of the stain and the amount of tissue remaining on the specimens. Craniums were dissected under the microscope.

Histological Evaluation of the Synchondroses at the Cranial Base

The bony bridges of a spheno-occipital synchondrosis and 2 anterior intraoccipital synchondroses were histologically examined after being stained with Alizarin red and were scored according to 4 separate appearances (non-bridge, 0 points; minimal bony bridge, 1 point; incomplete bony bridge, 2 points; complete bony bridge, 3 points). Therefore, the maximum bony bridge score was 9.

Measurements of Tissue Meclozine Concentration

After maternal ad libitum feeding of meclozine (0.4 g/kg diet) for 72 hours, ED 17.5 and PD 6.5 of C57BL/6 J mice were euthanized under deep anesthesia, and tissue concentrations of meclozine in the entire body were measured for evaluation of transmission of meclozine through the placenta and milk, respectively (Tanabe R&D Service Co.).

Statistical Analysis

Statistical analyses were carried out using the unpaired Student t-test. Data are expressed as the mean ± SD.

Results

Foramen Magnum Stenosis

We measured the area of the foramen magnum using reconstructed 3D images from micro-CT scanning in wild-type mice and Fgfr3<sup>ach</sup> mice at PD 17. The area of the foramen magnum was significantly decreased in Fgfr3<sup>ach</sup> mice (10.58 ± 0.42 mm<sup>2</sup>) compared with wild-type mice (12.97 ± 0.36 mm<sup>2</sup>) (p < 0.005) (Fig. 2).

Closure of Synchondroses

We next evaluated the effect of meclozine on synchondrosis closure at the cranial base in PD 4.5 mice. Sphen-occipital and anterior intraoccipital synchondroses were always cartilaginous in wild-type mice, whereas there were some bony bridges within these synchondroses in Fgfr3<sup>ach</sup> mice. We analyzed a total of 19 kinships, including 16 meclozine-treated Fgfr3<sup>ach</sup> mice and 19 untreated Fgfr3<sup>ach</sup> mice. Premature closure of occipital synchondroses was inhibited by meclozine treatment in some Fgfr3<sup>ach</sup> mice (Fig. 3). The average bony bridge score at the cranial base, however, was 7.053 ± 1.393 in untreated Fgfr3<sup>ach</sup> mice and 6.125 ± 2.029 in meclozine-treated Fgfr3<sup>ach</sup> mice, which was not statistically significant (p = 0.12) (Fig. 4).

Placental Transmission of Meclozine

We next examined the tissue concentrations of meclozine in ED 17.5 and PD 6.5 wild-type mice after maternal administration. Both the placental and breast milk transportability was confirmed by measurements of meclozine concentration in homogenized embryos and infants. The average tissue concentration of meclozine was 56.91 ± 20.05 ng/g in ED 17.5 mice and 508.88 ± 205.16 ng/g in PD 6.5 mice, indicating that placental transmission of meclozine was likely to be approximately 9 times less than that of breast milk transmission (Fig. 5).

Discussion

FMS is a devastating and life-threatening complication during early infancy in patients with ACH. In our previous study, we could not prevent FMS by administration of meclozine to 21- to 42-day-old Fgfr3<sup>ach</sup> mice, although the longitudinal bone lengths increased. Similarly, subcutaneous administration of BMN-111, in 7- to 17-day-old Fgfr3<sup>Y367C/+</sup> mice did not increase the sagittal and lateral diameters of the foramen magnum. In the present study, we found that premature closure of synchondroses at the cranial base began to develop in 4.5-day-old mutant mice, and FMS was evident in 17-day-old mutant mice. Maternal administration of an FGFR3-inhibiting drug, therefore, is indispensable to prevent premature closure of occipital synchondroses and FMS.
We previously demonstrated that maternal administration of meclozine from ED 14.5 to PD 4.5 increased longitudinal bone length in wild-type mice by 1.6% to 4.3% on PD 4.5 after adjusting litter size. The lengths of the ulna, femur, and tibia were significantly increased after meclozine treatment, while no statistically significant differences were seen in the lengths of the humerus and radius. Maternal administration from ED 14.5 to PD 4.5 showed less effect on longitudinal bone growth than oral administration to the growing pups from PD 21 to PD 42. In the present study, we could not show significant effects of maternal treatment of meclozine on premature closure of synchondroses in the cranial base at PD 4.5. Limited effects of prenatal treatment of meclozine on suppression of FGFR3 signaling may be due to its relatively low pla-
cental transmission. A higher dose of meclozine during the prenatal period would be necessary for the treatment of FMS in ACH, although teratogenicity and adverse effects of maternally administered meclozine would be of concern in clinical feasibility of this drug.

There are several limitations in the current study. We could not adjust the litter size of Fgfr3<sub>ach</sub> mice, which was different from mother to mother. In addition, the amount of breast milk intake might be different among mutant offspring, since some mutant mice were suffering from maternal neglect. The method of administration is another limitation. Pharmaceutical activity of meclozine within foods could be decreased over time. Further studies are needed to investigate the optimal dose of meclozine for the treatment of FMS in ACH.

Conclusions
The effect of maternal administration of meclozine on prevention of premature synchondrosis closure at the cranial base was not significant, probably due to low placental transmission of the drug.

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References

Disclosures
The authors report no conflict of interest concerning the materials or methods used in this study or the findings specified in this paper.

Author Contributions
Conception and design: Matsushita. Acquisition of data: Matsushita, Esaki. Analysis and interpretation of data: Matsushita, Esaki. Drafting the article: Kitoh, Matsushita, Ishiguro, Ohno. Revised submitted version of manuscript: Kitoh, Matsushita, Ishiguro, Ohno. Approved the final version of the manuscript on behalf of all authors: Kitoh. Statistical analysis: Matsushita. Administrative/technical/material support: Mishima, Esaki. Study supervision: Kitoh, Mishima, Ishiguro, Ohno.

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